

REGION TRACKING ALGORITHMS ON LASER SCANNING DEVICES APPLIED TO CELL TRAFFIC ANALYSIS

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Abstract

In vivo and *in situ* confocal images are often distorted by motion artifacts and soft tissue deformations. To measure small amplitude phenomena on this type of images, we have to compensate for those artifacts. We present in this paper a Region Of Interest (ROI) tracking algorithm specialized for confocal imaging using a scanning device. Two different algorithms are presented: one based on the motion artifacts, and one based on affine registration. One typical application of this tool is developed: the blood velocity estimation inside a capillary on a moving organ. These first results show that the method permits accurate estimations of blood cell velocities even in presence of motion artifacts.

In vivo Imaging and Motion Artifacts

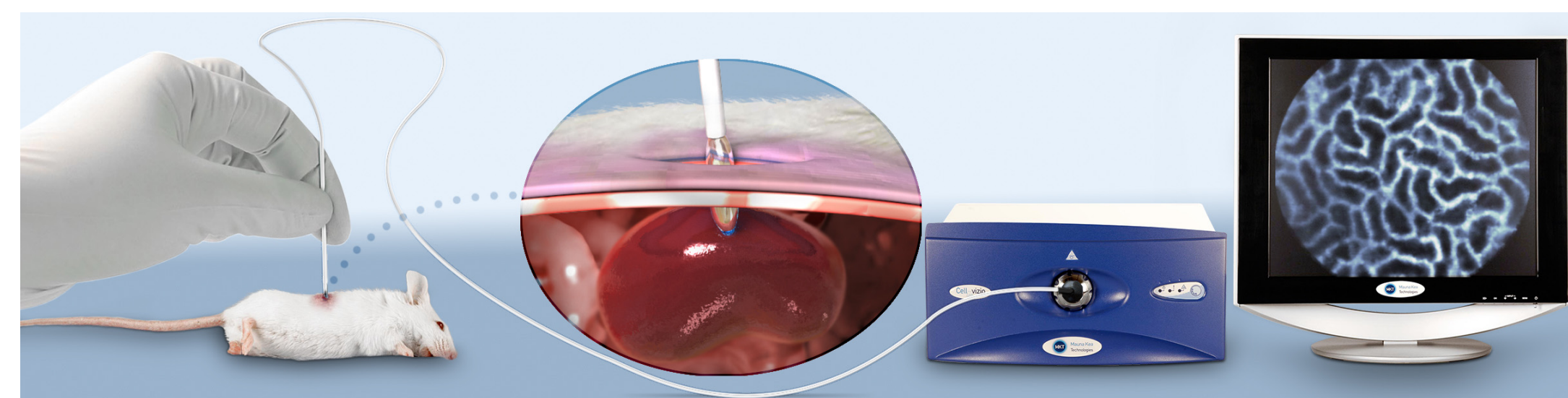


Fig. 1. Typical minimally invasive experimental setup with Cellvizio®. The kidney is accessed through a buttonhole.

Cellvizio® is a complete imaging system distributed by in North America, Europe and Japan under the name Leica FCM-1000. It is based on a fibered technology for fluorescence confocal imaging of the living animal. It acquires microscopic resolution sequences, displays them in real-time and enables live measurements.

- *In vivo* confocal microscopy
- Create motion artifacts due to tissue or objective motion
- Classical image stabilization and registration technique are not directly applicable

Still, there is a growing need for quantification on specific moving organs: liver, bladder, heart...

- Pharmacokinetics parameters of molecules, Changing morphology of micro-structures, Bio distribution parameters...



Fig. 2. Slit-scan photo by J.H. Lartigue

Region Tracking

Motion Compensation Algorithm

- Estimation of translation using 2D cross-correlation
- Estimation of the velocity: $\tilde{\eta} = [\tilde{\eta}^x, \tilde{\eta}^y]$
- Computation of the distortion transformation: v_k

$$\begin{cases} x_d = x + (t(x) - t(0))\tilde{\eta}^x = x + (y/v_y)\tilde{\eta}^x = x + \eta^x y, \\ y_d = y + (t(y) - t(0))\tilde{\eta}^y = y + (y/v_y)\tilde{\eta}^y = (1 + \eta^y)y. \end{cases}$$
- Optimization of the rigid transformation: r_k

The resulting transformation between frames j and k is:

$$f_{j,k}(p) = v_j^{-1} \circ r_j^{-1} \circ r_k \circ v_k = v_j^{-1} \circ r_{j,k} \circ v_k$$

Affine Registration Algorithm

- Initialize translation using 2D cross-correlation
- Optimize Affine Transformation T_k^a using SSD criterion

$$\sum_p \|I(p) - I_k(T_k^a(p))\|^2$$

Performance Comparison

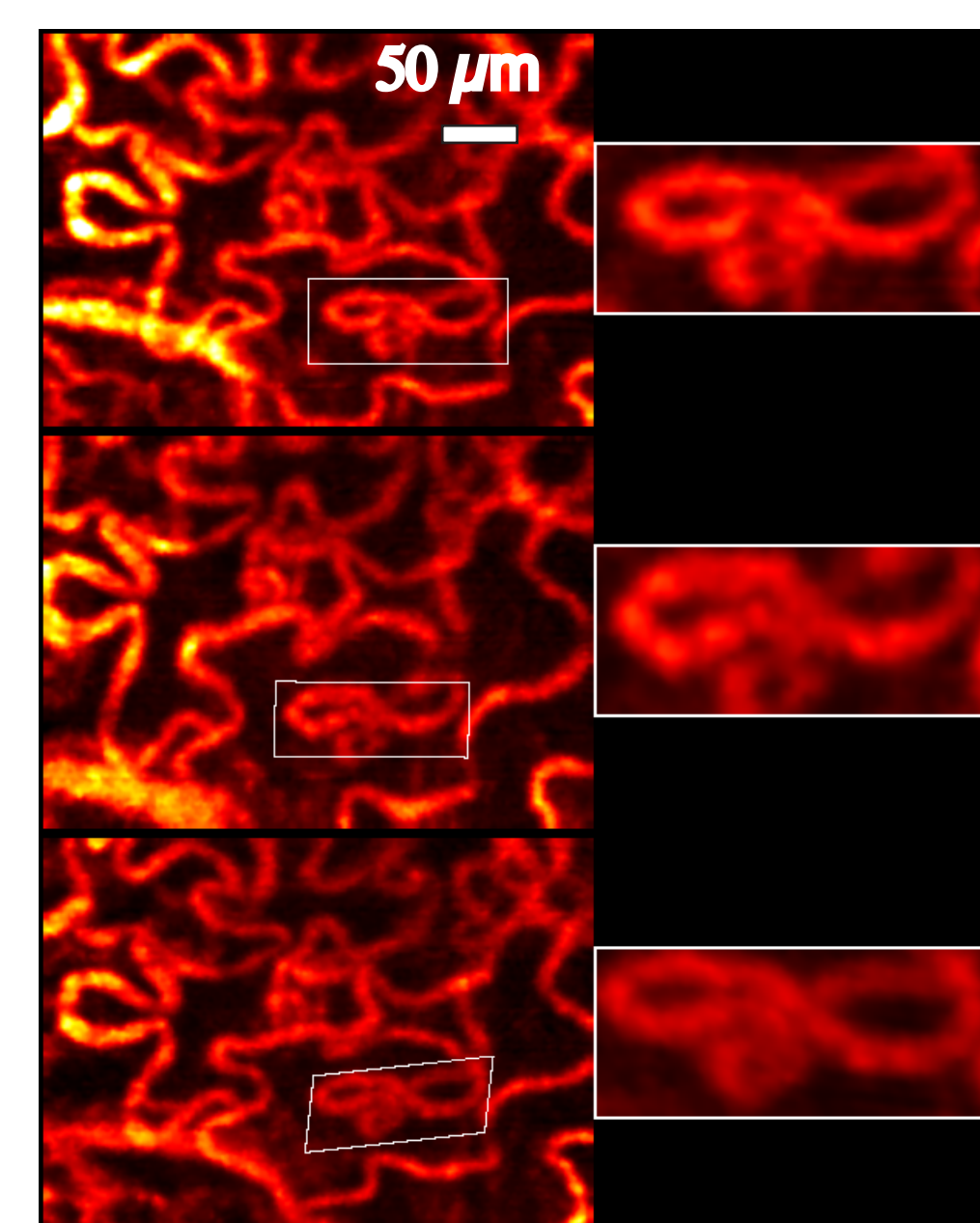


Fig. 3. Motion Compensation Algorithm

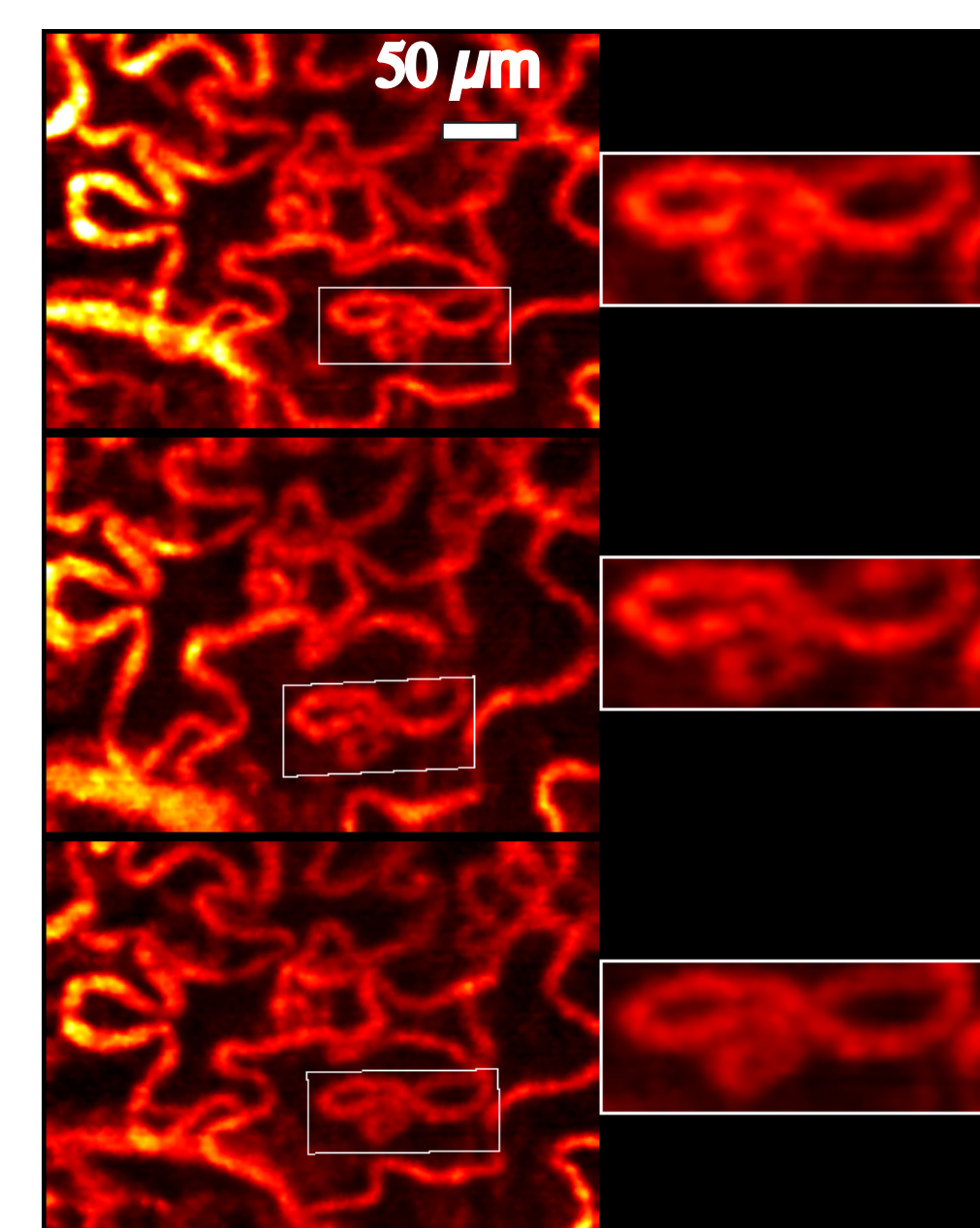


Fig. 4. Affine Registration Algorithm

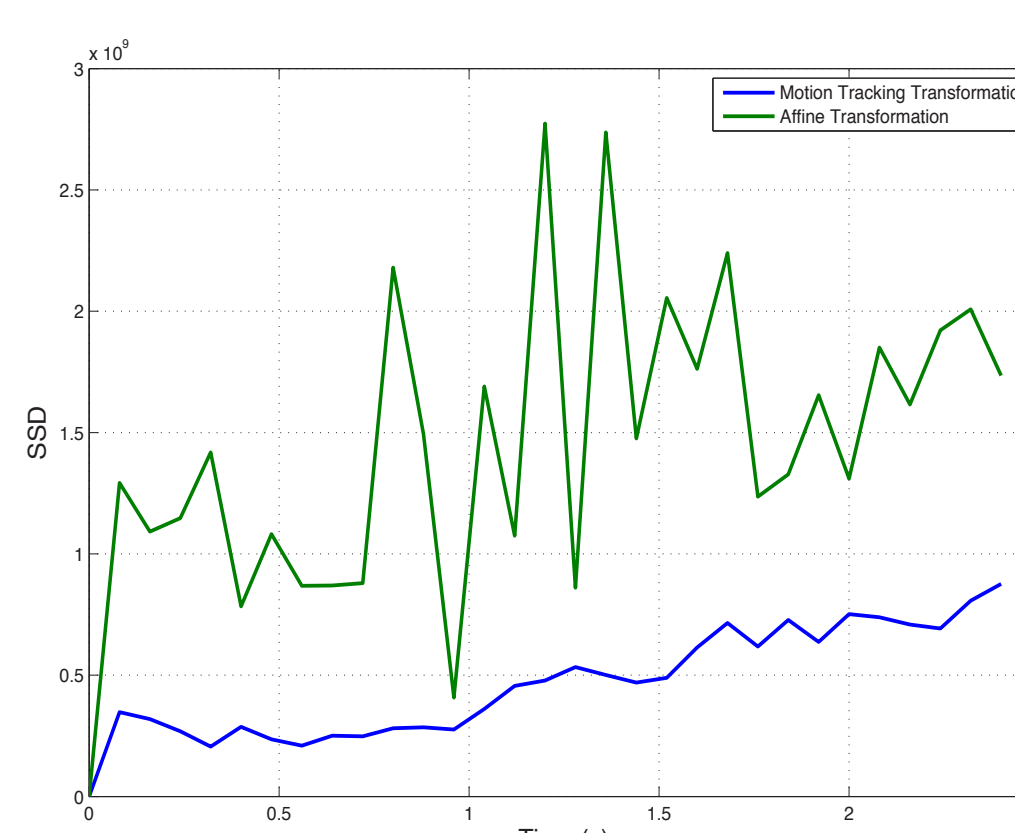


Fig. 5. SSD measurement on final result

Motion Compensation
+ physic (model) based
+ more stable
+ faster

Affine Registration
+ more generic
+ if good stability, can be more accurate

Application To Cell Traffic Analysis

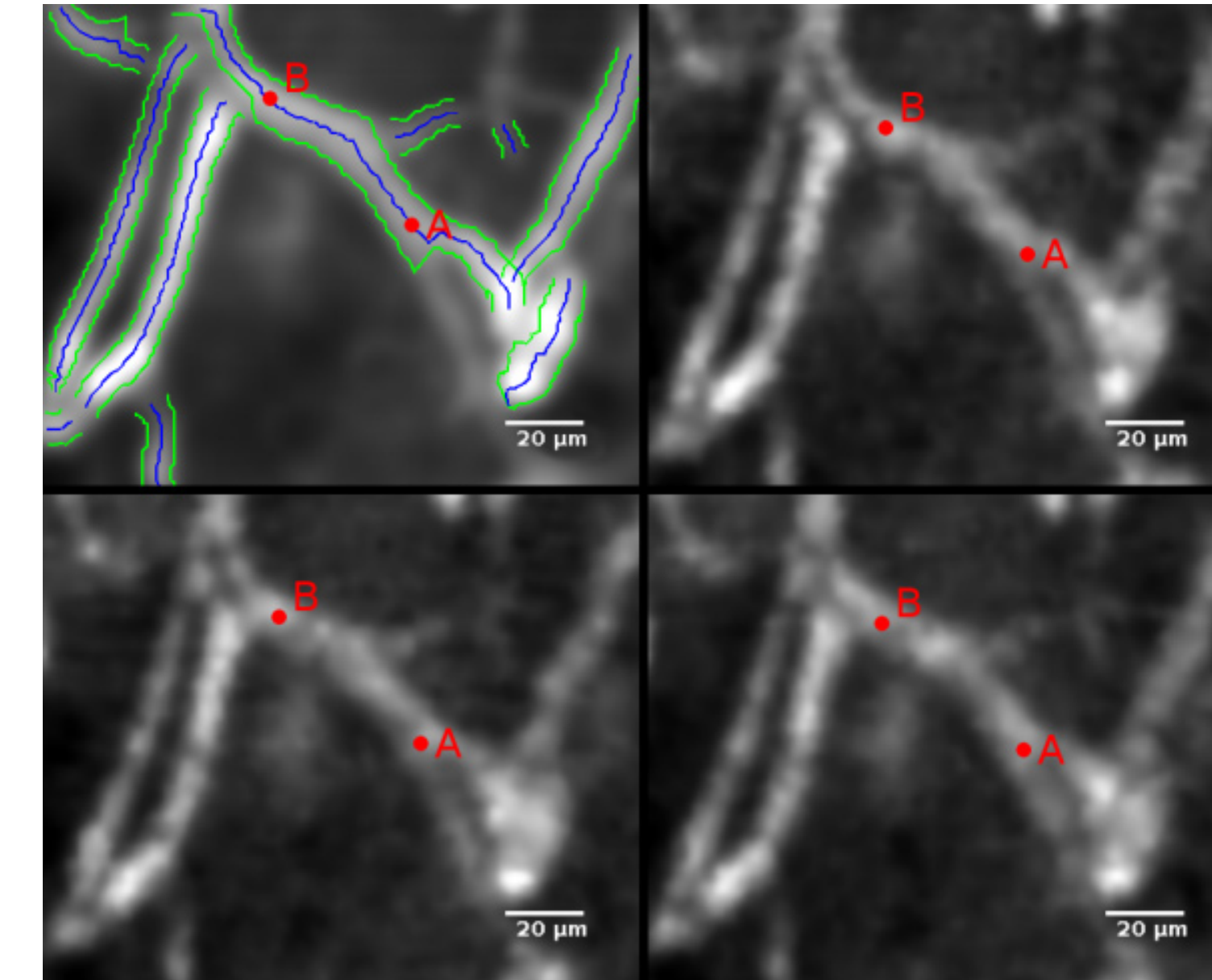


Fig. 6. Vessel Detection on mean image, and spatio-temporal analysis of blood flow along medial axis AB.

- In vivo imaging of a tumoral skin Xenograft on a mouse
- Manual selection of a Region Of Interest
- Automatic tracking of the ROI
- Vessel Detection on the temporal mean ROI (Krissian et. al., 2000)
- Spatio-temporal analysis by cross correlation

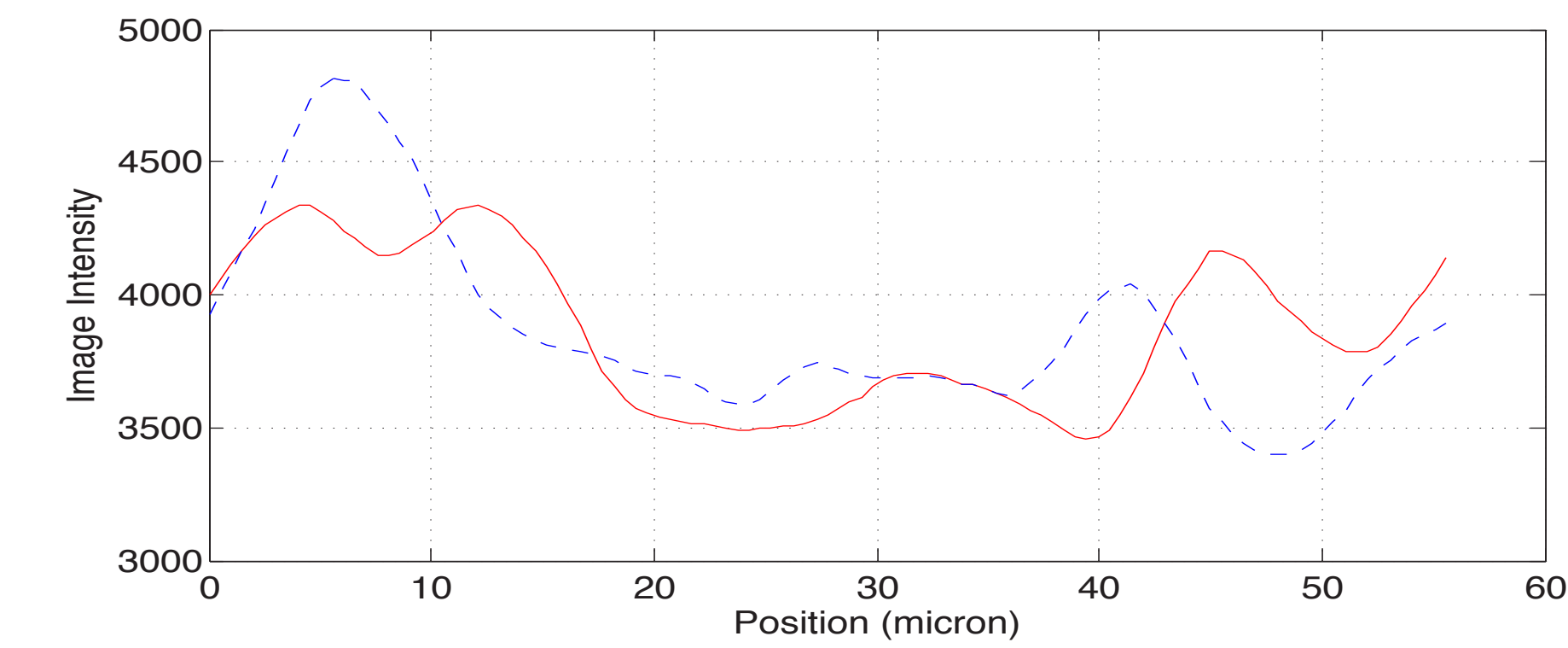


Fig. 7. A-B lines from medial axis detection on two contiguous frames.

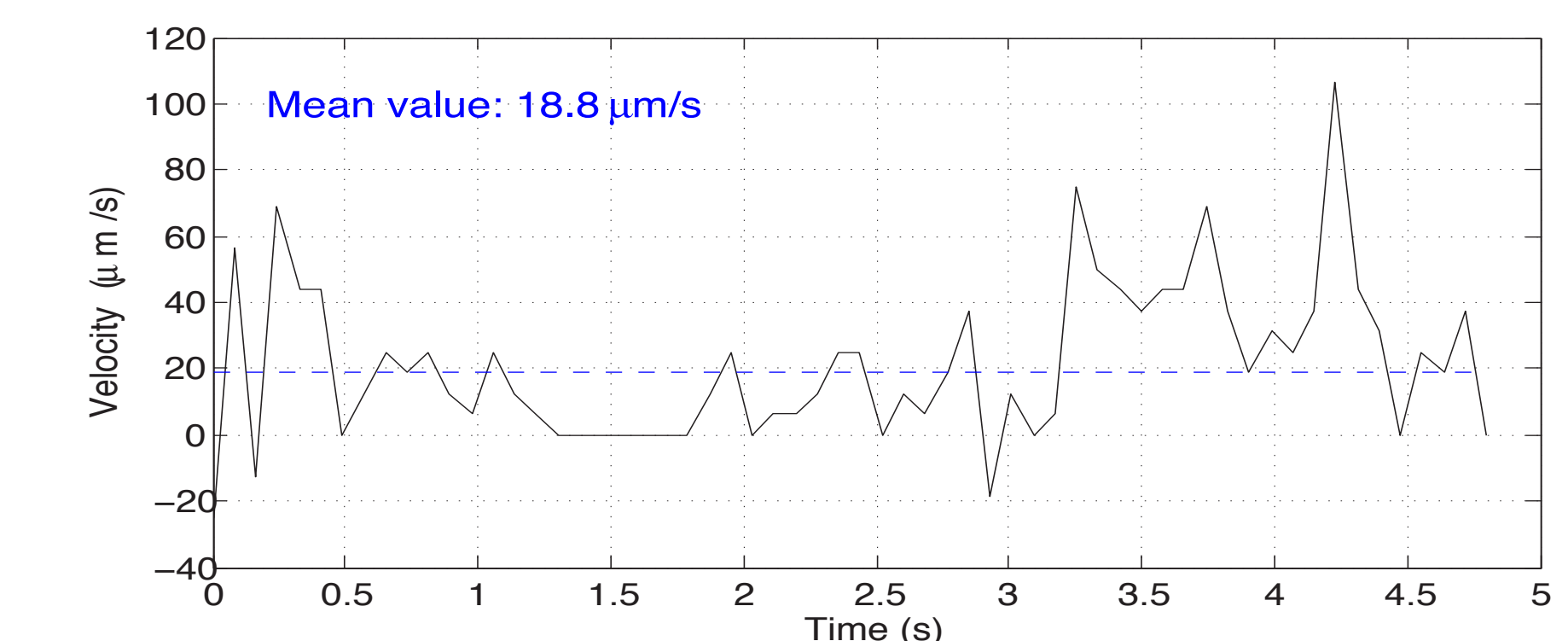


Fig. 8. Velocity estimation at 12Hz frame rate after motion tracking. The blue line is the mean velocity.

Contributions and Conclusion

In vivo and *in situ* microscopic imaging bring new constraints to image quantification: constant motion and large amount of data.

Our contribution is to bridge the gap between *in vivo in situ* confocal microscopy and standard confocal microscopy when dealing with morphological quantification:

- Bring stability as a preprocessing step,
- Motion tracking is based on physical model (motion compensation algorithm), or includes more generic transformation (affine)
- In the near future: stabilization should be real-time

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